



Comparison of nuclear abnormalities in *Astyanax bifasciatus* Cuvier, 1819 (Teleostei: Characidae) of two sections of rivers from the middle Iguaçu

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ABSTRACT. Most changes in water bodies are result of human activities that have the potential to undermine the environmental integrity of aquatic ecosystems. Changes in genetic material can be evidenced by the frequency of nuclear abnormalities in fish blood cells, in response to genotoxic agents even at low concentrations. Thus, we aimed at comparing the frequencies of nuclear abnormalities of fish kept in acclimation, with fish collected in Timbó river (Santa Cruz do Timbó, Santa Catarina State), under low anthropogenic interference, and fish collected in Iguaçu river (União da Vitória, Paraná State), a polluted river. The highest frequency of changes in nuclear morphology of fish erythrocytes was found in the urbanized area around the Iguaçu river, while, the fish collected in areas with preserved riparian forest surrounding the Timbó river and acclimated fish showed no morphological changes. Our results suggest that genotoxic compounds in the Iguaçu river are acting as stressors to aquatic communities, especially fish. In addition, the absence of nuclear abnormalities in fish from the Timbó river suggests that rivers without significant environmental changes can serve as reference point for comparative studies of genetic modifications for the species studied.

Keywords: micronucleus; ecotoxicology; mutagenesis; pollution.

Comparação de anormalidades nucleares em *Astyanax bifasciatus* Cuvier, 1819 (Teleostei: Characidae) de dois trechos de rios do médio Iguaçu

RESUMO. A maioria das alterações dos corpos hídricos é resultante das atividades antrópicas que são prejudiciais à integridade ambiental dos ecossistemas aquáticos. Alterações no material genético podem ser evidenciadas pela frequência de anormalidades nucleares em peixes, em resposta a agentes genotóxicos, mesmo em baixas concentrações. Dessa forma, o estudo consistiu em comparar as frequências de anormalidades nucleares de peixes mantidos em aclimação, com peixes coletados no rio Timbó (Santa Cruz do Timbó, Estado de Santa Catarina), local com pouca interferência antropogênica e, peixes coletados no rio Iguaçu (União da Vitória, Estado do Paraná), um rio poluído. As maiores frequências de alterações na morfologia nuclear dos eritrócitos dos peixes foram visualizadas na área urbanizada ao redor do rio Iguaçu, quando comparados com os peixes coletados em área com mata ciliar preservada, no rio Timbó, e os peixes aclimatados, que não apresentaram alterações morfológicas. Esses resultados sugerem que há compostos genotóxicos no trecho médio do rio Iguaçu que são estressores para os peixes. Além disso, a não observação de anormalidades nucleares no rio Timbó sugere que rios sem alterações ambientais podem servir como ponto de referência para estudos comparativos de modificações genéticas para a espécie estudada.

Palavras-chave: micronúcleo; ecotoxicologia; mutagênese; poluição.

Introduction

Like any aquatic environment, the basin of the Iguaçu river is exposed to pollution processes caused by the huge variety and quantity of chemicals (Livingstone, 1998), and presents pollution evidence from industrial waste from the manufacture of paper, fertilizers and waste from agricultural practice (Bueno-Krawczyk et al., 2015). These toxic

substances released into the environment may interact with biota, causing changes (Arias et al., 2007) that can have a genotoxic potential and/or mutagenic and clastogenic effects (Zenkner, Soares, Prá, Köhler, & Rieger, 2011), favoring the formation of micronucleus and nuclear abnormalities as they can be absorbed and stored in the animal body (Benites, Doncato, Minho, & Perazzo, 2014).

Different types of nuclear lesions have been described and classified as “Blebbled”, “Notched”, “Lobed”, “Vacuolated” and “Micronucleus” (Carrasco, Tilbury, & Myers, 1990). These abnormalities are used as indicators of genotoxic damage (Pacheco & Santos, 1998; Aylon & Garcia-Vazquez, 2000).

Therefore, environmental stress can be detected early using bio-indicators (Brito & Luz, 2015), among which fish species are recommended (Cotelle & Ferrard, 1999) since they play an important ecological role in the food chains, transferring energy to other trophic levels (Cort & Ghisi, 2014). Moreover, these organisms have genetic structure similar to mammals (Barbazuk et al., 2000; Lieschke & Currie, 2007) responding similarly to environmental contamination.

Species of the genus *Astyanax* have been used as biological models for different studies because they have dynamic life cycles with a high reproductive potential (Zenkner et al., 2011), having capacity of absorbing quickly compounds which are directly added into the water and accumulate them in different tissues (Silva, 2014), as well as endemic species to the Iguaçú river basin, showing thus high abundance (Garavello & Sampaio, 2010). Studies using this genus as bioindicators in the environmental integrity analysis have been widely spread (Zenkner et al., 2011).

A tool used for genotoxicity check on a species is the piscine micronucleus (Hooftman & Raat, 1982). Micronuclei are whole chromosomes or partials that are not incorporated into the nucleus of the daughter cell during cell division and which appear as a small round dark structure, identical in appearance to the cell nucleus (Al-Sabti & Metcalfe, 1995); they may reflect exposure to agents with clastogenic (chromosome breakage, when DNA is the target of the chemical agent) or aneugenic (effect on the number of chromosomes in most cases, the chemical agent does not target the DNA) action mode (Albertini et al., 2000).

Studies correlating the alterations in fish health due to water pollution in the Iguaçú river basin are incipient and the existing ones demonstrate the presence of drugs, such as naproxen, salicylic acid, acetylsalicylic acid (Ide, Osawa, Marcante, Pereira, & Azevedo, 2017), phosphorus (Sodré, Schnitzler, Scheffer, & Grassi, 2012) and caffeine in the Middle Iguaçú region (Bueno-Krawczyk et al., 2015), commonly used as indicator of anthropic activity in natural environments, related to the discharge of untreated domestic sewage (Seiler, Zaugg, Thomas, & Howcroft, 1999).

Therefore, in order to verify the response pattern generated by the analysis of micronucleus in fish erythrocytes and the response of this biomarker to

water pollution, we compared the frequencies of nuclear abnormalities in fish of the species *Astyanax bifasciatus* collected in the Timbó river with fish collected in the Iguaçú river, and fish kept in acclimation for 60 days.

Material and methods

Study area and fish sampling

We defined two sampling sites. In the Iguaçú river (26°13'41.7"S and 051°05'59.6"W), fish (n=15) (IBAMA license - SISBIO 40229-1) were collected in a point of the river localized in União da Vitória, Paraná State, under strong anthropogenic interference, with many surrounding houses and absence of riparian vegetation (Figure 1 - P1). In the Timbó river (26°25'45"S e 50° 50' 46.1" W), fish were collected in a point of the river localized in Santa Cruz do Timbó, Santa Catarina State, under little anthropic interference and preserved riparian vegetation (Figure 1 - P2). In this sampling, 20 fish were collected (IBAMA license - SISBIO 42950-1), of which 10 were analyzed soon after collection and, the remainder (n = 10) were transported to the Pisciculture of the *Universidade Estadual do Paraná* (UNESPAR), where they remained in acclimation conditions with filtered water, available oxygen and daily feeding for 60 days.

Water chemical analysis

For metal analysis, we collected surface water of the Iguaçú river and stored in 500 mL polyethylene bottles. The sample was analyzed, after centrifuging, by inductively coupled plasma optical emission (ICP OES) for large and selected oligoelements. The spectrophotometry was done by atomic absorption according to American Public Health Association (APHA, 2012). This analysis of heavy metals was performed only for the sample from the Iguaçú river due to its known history of pollution (*Instituto Brasileiro de Geografia e Estatística* [IBGE], 2012).

Samples

In laboratory, fish were anesthetized with clove oil diluted in water (1.5 $\mu\text{L L}^{-1}$). For the piscine micronucleus test, blood samples were obtained by caudal puncture with heparinized syringes. For each specimen, blood was drawn and one thin-layer slide was prepared using a drop of blood. Blood smears were air-dried for 24 hours, fixed in absolute ethanol for 10 min., and then stained with 10% Giemsa solution for 10 min. Afterwards, they were washed with running water. A total of 2000 cells per slide were analyzed at 1000 \times magnification on an optical microscope. The Micronucleus test was performed according to Hooftman and Raat (1982). For the analysis of erythrocyte nuclear abnormalities (ENA), we used the classifications proposed by Carrasco et al. (1990).

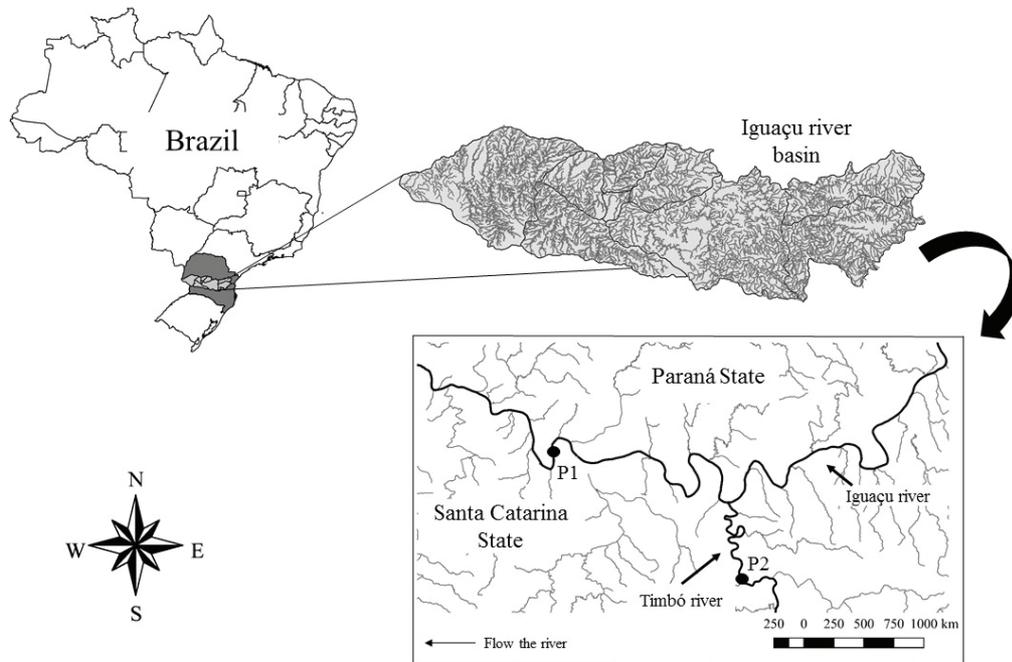


Figure 1. Geographical localization of sampling sites: Iguazu river, Paraná (P1) and Timbó river, Santa Catarina (P2).

Data analysis

Since our data were independent samples and did not reach normality and homoscedasticity assumptions for analysis of variance, we used the Kruskal-Wallis non-parametric equivalent, followed by Dunn's post-hoc test, to identify significant differences between the analyzed rivers and the acclimation group. We used Vegan (Oksanen et al., 2016) and the Dunn Test (Dinno & Dinno, 2017) statistical packages in software R (Core Team 2018). Statistical significance was accepted at $p < 0.05$.

Results

We did not observe the occurrence of cell and micronucleus abnormalities in the fish collected in the Timbó river and the fish left in acclimation. In Iguazu river, fish showed high frequencies of nuclear abnormalities and frequency (1.15%) of micronucleus (0.35%).

In addition to micronucleus, we recorded three types of alterations in the nuclear morphology of blood cells of *Astyanax bifasciatus*: Lobed, Blebbed and Notched (Figure 2).

When we compared the nuclear abnormalities between the fish collected in the Iguazu river, Timbó and in acclimation (Table 1), we observed that the fish of the Iguazu river presented significant

differences in the frequency of nuclear abnormalities for Notched, Blebbed and micronucleus when compared with the Timbó river and Acclimation group, whereas for Lobed, no significant differences were detected between these groups (Table 2 and Figure 3).

Table 1. Kruskal-Wallis test for differences in the frequency of Lobed, Blebbed, Notched and Micronucleus between Iguazu river, Timbó river and acclimation group. Bold p-values indicate significance at $p < 0.05$ (df = degrees of freedom; X^2 =Chi-Squared).

Kruskal-Wallis test	df	X^2	p-value
Lobed	2	5.833	0.054
Blebbed	2	7.504	0.023
Notched	2	9.302	0.009
Micronucleus	2	7.512	0.009

Table 2. Dunn's post-hoc test for differences in the frequency of Blebbed, Notched and Micronucleus between Iguazu river, Timbó river and acclimation group. Bold p-values indicate significance at $p < 0.05$.

Dunn's test	p-value
Blebbed	
Iguazu x Timbó	0.032
Iguazu x Acclimation	0.032
Timbó x Acclimation	1.000
Notched	
Iguazu x Timbó	0.016
Iguazu x Acclimation	0.016
Timbó x Acclimation	1.000
Micronucleus	
Iguazu x Timbó	0.032
Iguazu x Acclimation	0.032
Timbó x Acclimation	1.000

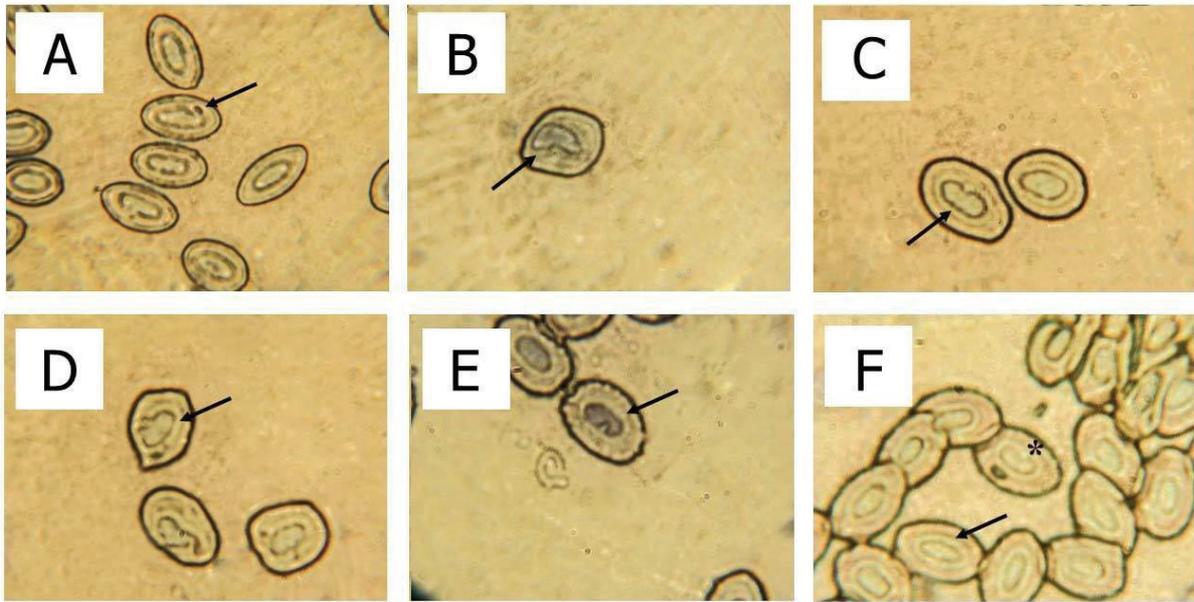


Figure 2. Micronucleus and nuclear abnormalities (stained with Giemsa) found in erythrocytes of *Astyanax bifasciatus* from Iguaçu river. (A) Micronucleus (40X); (B and C) Lobed (40X); (D) Blebbed (40X) (E) Notched (40X) (F) normal cell (arrow) (40X) and Notched (*).

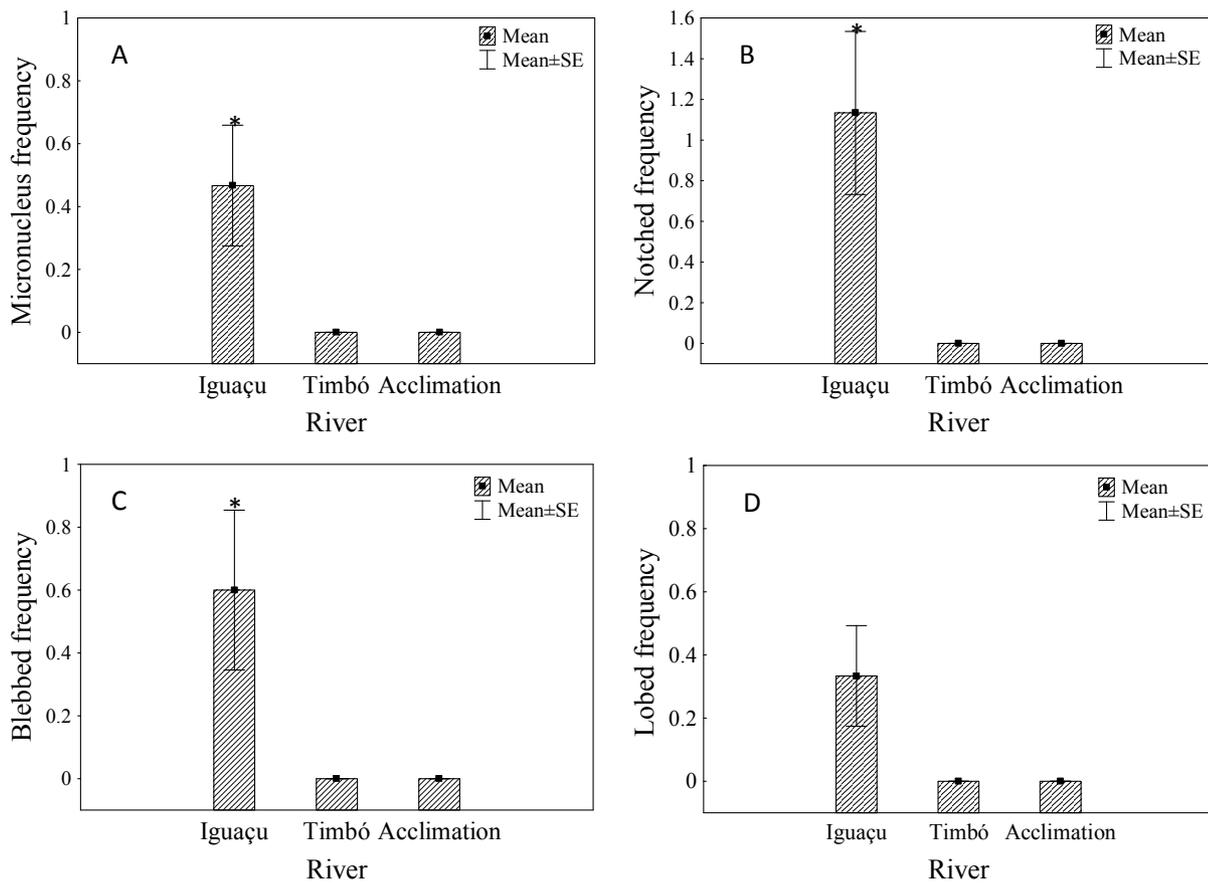


Figure 3. Frequency of micronucleus nuclear abnormalities in *Astyanax bifasciatus* from Iguaçu river, Timbó river and in acclimation. * Significant difference ($p < 0.05$). (A) Micronucleus; (B) Notched; (C) Blebbed; (D) Lobed.

Of the metals tested in the Iguaçú river, the concentrations of aluminum (0.3 mg L^{-1}) and of lead (0.02 mg L^{-1}) exceeded the maximum value (0.1 and 0.01 mg L^{-1} respectively) allowed by *Conselho Nacional do Meio Ambiente* (CONAMA, 2005) resolution 357/05 for class 3 waters (Table 3). Other metals did not show values above the established by the Resolution.

Table 3. Values found in the characterization of heavy metals in the sampling site in the Iguaçú river. Bold numbers represent the values above the acceptable.

Analyzed metals	Maximum values allowed (mg L^{-1})	Values found in the Iguaçú river (mg L^{-1})
Aluminum	0.1	0.369
Barium	0.7	0.030
Cadmium	0.001	-0.002
Cobalt	0.05	0.035
Chrome	0.05	0.137
Copper	0.009	0.002
Iron	0.3	-0.042
Magnesium	0.1	-0.000
Manganese	0.1	-0.000
Nickel	0.025	-0.005
Lead	0.01	0.028
Vanadium	0.1	0.050
Zinc	0.18	-0.002

Discussion

Fish of the Timbó river showed no cell nucleus morphological change, which may be due to the riparian forest of their surroundings that has important role in preservation of aquatic ecosystems, since it retains residues provided by leaching, controls fluvial dynamics besides absorbing pollutants reducing the ecotoxicological effects on aquatic biota, including fish (Peterjohn & Correl, 1984; Tabacchi et al., 1998; Lobón-Cerviá, Hess, Melack, & Araújo-Lima, 2015).

The opposite was observed in the Iguacu river, where fish collected presented high frequencies of nuclear abnormalities, including the occurrence of micronucleus. This may be due to the absence of riparian vegetation and the high degree of human disturbance, mining, dumping of waste from the pulp industry and disposal of waste from sewage treatment. These abnormalities and the presence of micronucleus may be related to a chronic exposure of the water ecosystem, since these activities generally cause degradation of water resources, affect the aquatic biota in addition to altering the physical and chemical characteristics of the water (Helena, Vega, Barrado, Pardo, & Fernández, 1999) causing impacts on the water body (Silva & Nepomuceno, 2010; Ribeiro et al., 2013; Cort & Ghisi, 2014). The effects of pollution cause impacts on the water body (Silva & Nepomuceno, 2010; Ribeiro et al., 2013; Cort & Ghisi, 2014), which may affect the structure

of fish populations (Bifi, Baumgartner, Baumgartner, Frana, & Debona, 2006) and, cause long-term, genetic changes in species.

Hemachandra and Pathiratne (2016) warn of the effects of industrialization on the ecological integrity of aquatic ecosystems. Indeed, the Iguaçú river receives large discharges of domestic and industrial effluents without treatment and, some authors observed an increase in the number of nuclear alterations of the Lobed and Blebbed types and micronucleus formation in fish exposed to concentrations of industrial effluents from textile factories (Çavas & Ergene-Gozukara, 2003). Besides that, exposure to drugs can cause negative effects on aquatic organisms (Ribas, Zampronio, & Assis, 2015), even at low concentrations (Guiloski, Ribas, Pereira, Perbiche, & Assis, 2015). For example, Ragunetti et al. (2011) observed that the frequency of micronucleus for the species *Oreochromis niloticus* increased proportionally with time of exposure to ibuprofen.

Furthermore, in our study, the presence of aluminum three times above that allowed by Brazilian legislation, found in the Iguaçú river, is worrisome since this metal causes toxic effects to biota, especially when exposure occurs over a prolonged period (Gensemer & Playle, 1999). Some studies have shown the aluminum ability to promote the formation of micronucleus and nuclear morphological changes in fish. García-Medina et al. (2013), when determining the cytotoxic and genotoxic effects of common carp (*Cyprinus carpio*) erythrocyte exposure to aluminum, found that the aluminum concentrations tested produced higher oxidative stress and induced higher frequencies of micronucleus in the species. Likewise, Galindo, Troilo, Cólus, Martínez, and Sofia, (2010), observed the formation of nuclear abnormalities in erythrocytes of *Prochilodus lineatus*, when exposed to different concentrations of aluminum. Similarly, lead also causes changes once it is a quite toxic metal (Türkmen, Türkmen, Tepe, Töre, & Ates, 2008) capable of altering vital fish structures, causing edema and cerebral hemorrhages (Adeyemo, 2008), hepatic and hematological diseases in the organisms (Ates, Orun, Talas, Durmaz, & Yilmaz, 2008), and morphologically alter the cellular nucleus of fish, as reported by Ferraro, Fenocchio, Mantovani, Ribeiro, and Cestari (2004). Moreover, fish are bio-concentrator organisms and many contaminants, even that at low concentrations, can affect their physiology and survivability (Grisolia et al., 2009).

We verified that *Astyanax bifasciatus* is sensitive to pollution. The same species was used as a chemical

and genetic bioindicator in the middle section of the Iguaçú river by Bueno-Krawczyk et al. (2015), who observed changes in fish health conditions probably related to the presence of heavy metals and caffeine in the environment. The *Astyanax* genus has been used as a bioindicator because of its sensitivity to anthropogenic actions, such as agricultural conditions and wastewater, presenting cellular stress (Costa-Silva et al., 2015; Matozo, Turek, & Noletto, 2015) and higher frequencies of nuclear abnormalities, including the micronucleus formation (Dalzochio et al., 2018).

Conclusion

We concluded that Iguaçú river contains contaminants that cause genotoxicity effects in fish. Fish exposed to water of this river showed nuclear abnormalities and micronuclei in the blood cells while fish from Timbó river and in the acclimation group did not. The results emphasized the need for discussion about Iguaçú river conservation and regulation of the inputs of contaminants into water bodies. Timbó river is a water body without significant environmental changes and can serve as reference point for comparative studies of genetic modifications for the studied species.

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References

- American Public Health Association [APHA]. (2012). *Standard Methods for the Examination of Water and Wastewater* (22th ed.).
- Adeyemo, O. K. (2008). Histological alterations observed in the gills and ovaries of *Clarias gariepinus* exposed to environmentally relevant lead concentrations. *Journal of Environmental Health*, 70(9), 48-51.
- Albertini, R. J., Anderson, D., Douglas, G. R., Hagmar, K. H., Merlo, F., Natarajan, ..., Aitio, A. (2000). IPCS guidelines for the monitoring of genotoxic effects of carcinogens in humans. *Mutation Research*, 463(2), 111-172. doi: 10.1016/S1383-5742(00)00049-1
- Al-Sabti, K., & Metcalfe, C. D. (1995). Fish micronuclei for assessing genotoxicity in water. *Mutation Research/Genetic Toxicology*, 343(2-3), 121-135. doi:10.1016/0165-1218(95)90078-0
- Arias, A. R. L., Buss, D. F., Albuquerque, C. D., Inácio, A. F., Freire, M. M., Egler, M., ... Baptista D. F. (2007). Utilização de bioindicadores na avaliação de impacto e no monitoramento da contaminação de rios e córregos por agrotóxicos. *Ciência & Saúde Coletiva*, 12(1), 61-72. doi: 10.1590/S1413-81232007000100011
- Ates, B., Orun, I., Talas, Z. S., Durmaz, G., & Yilmaz, I. (2008). Effects of sodium selenite on some biochemical and hematological parameters of rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792) exposed to Pb²⁺ and Cu²⁺. *Fish Physiology Biochemistry*, 34(1), 53-59. doi: 10.1007/s10695-007-9146-5
- Aylon, F., & Garcia-Vazquez, E. (2000). Induction of micronuclei and other nuclear abnormalities in European minnow *Phoxinus phoxinus* and mollie *Poecilia latipinna*: an assessment of the fish micronucleus test. *Mutation Research - Genetic Toxicology and Environmental*, 467(2), 177-186. doi: 10.1016/S1383-5718(00)00033-4
- Barbazuk, W. B., Korf, I., Kadavi, C., Heyen, J., Tate, S., Wun, E., ... Johnson S. L. (2000). The syntenic relationship of the Zebrafish and human genomes. *Genome Research*, 10(9), 1351-1358. doi: 10.1101/gr.144700
- Benites, L. M., Doncato, K. B., Minho, T. S., & Perazzo, G. X. (2014). A avaliação do potencial mutagênico de cobre da água do rio Uruguai. *Ciência e Natura*, 36(2), 107-113. doi: 10.5902/2179460X13610
- Bifi, A. G., Baumgartner, D., Baumgartner, G., Frana, V. A., & Debona, T. (2006). Composição específica e abundância da ictiofauna do rio dos Padres, bacia do Rio Iguaçú, Brasil. *Acta Scientiarum - Biological Sciences*, 28(3), 203-211. doi: 10.4025/actascibiolsci.v28i3.193
- Brito, L. O., & Luz, L. Y. D. (2015). Avaliação e monitoramento da qualidade das águas: Usando análises moleculares. *Gesta*, 3(2), 76-90. doi: 10.17565/gesta.v3i2.13856
- Bueno-krawczyk, A. C. D., Guiloski, I. C., Piancini, L. D. S., Azevedo, J. C., Ramsdorf, W. A., Ide, A. H., ... Silva de Assis, H. C. (2015). Multibiomarker in fish to evaluate a river used to water public supply. *Chemosphere*, 135, 257-264. doi: 10.1016/j.chemosphere.2015.04.064
- Carrasco, K. R., Tilbury, K. L., & Myers, M. S. (1990). Assessment of the piscine micronucleus test as an in situ biological indicator of chemical contaminant effects. *Canadian Journal of Fisheries and Aquatic Sciences*, 47(11), 2123-2136. doi: 10.1139/f90-237
- Conselho Nacional do Meio Ambiente [CONAMA]. (2005). Resolução nº 357. Dispõe sobre a classificação dos corpos de água e diretrizes ambientais para o seu enquadramento, bem como estabelece as condições e padrões de lançamento de efluentes, e dá outras providências. *Diário Oficial da União*, 17 de Março de 2005. Seção 1, p. 58063.2005.
- Cort, C. C. W. D., & Ghisi, N. C. (2014). Uso de alterações morfológicas nucleares em *Astyanax* spp. para avaliação da contaminação aquática. *O Mundo da Saúde*, 38(1), 31-39. doi: 10.15343/0104-7809.20143 801031039
- Costa-Silva, D. G., Nunes, M. E. M., Wallau, G. L., Martins, I. K., Zemolin, A. P. P., Cruz, L. C., ...

- Franco, J. L. (2015). Oxidative stress markers in fish (*Astyanax* sp. and *Danio rerio*) exposed to urban and agricultural effluents in the Brazilian Pampa biome. *Environmental Science and Pollution Research*, 22(20), 15526-15535. doi: 10.1007/s11356-015-4737-7
- Cotelle, S., & Ferrard, J. F. (1999). Comet assay in genetic ecotoxicology: a review. *Environmental and Molecular Mutagenesis*, 34(4), 246-255.
- Çavas, T., & Ergene-Gozukara, S. (2003). Micronuclei, nuclear lesions and interphase silver-stained nucleolar organizer regions (AgNORs) as cyto-genotoxicity indicators in *Oreochromis niloticus* exposed to textile mill effluent. *Mutation Research*, 538(1), 81-91. doi: 10.1016/S1383-5718(03)00091-3
- Dalzochio, T., Rodrigues, G. Z. P., Simões, L. A. R., Souza, M. S., Petry, I. E., Andriguetti N. B., ... Gehlen, G. (2018). In situ monitoring of the Sinos River, southern Brazil: water quality parameters, biomarkers, and metal bioaccumulation in fish. *Environmental Science and Pollution Research*, 25(1), 1-16. doi: 10.1007/s11356-018-1244-7
- Dinno, A., & Dinno, M. A. (2017). *Dunn's Test of Multiple Comparisons Using Rank Sums*. R package version, 1.3.4. Retrieved on Nov. 12, 2017 from <https://cran.r-project.org/web/packages/dunn.test/dunn.test.pdf>
- Ferraro, M. V. M., Fenocchio, A. S., Mantovani, M. S., Ribeiro, C. D. O., & Cestari, M. M. (2004). Mutagenic effects of tributyltin and inorganic lead (Pb II) on the fish *H. malabaricus* as evaluated using the comet assay and the piscine micronucleus and chromosome aberration tests. *Genetics and Molecular Biology*, 27(1), 103-107. doi: 10.1590/S1415-47572004000100017
- Galindo, B. A., Troilo, G., Cólus, I. M. S., Martinez, C. B., & Sofia, S. H. (2010). Genotoxic effects of aluminum on the neotropical fish *Prochilodus lineatus*. *Water, Air, & Soil Pollution*, 212(1-4), 419-428. doi: 10.1007/s11270-010-0357-5
- Garavello, J. C., & Sampaio, F. A. A. (2010). Five new species of genus *Astyanax* Baird & Girard, 1854 from Rio Iguaçú, Paraná, Brazil (Ostariophysi, Characiformes, Characidae). *Brazilian Journal of Biology*, 70(3), 847-865. doi: 10.1590/S1519-69842010000400016
- García-Medina, S., Núñez-Betancourt, J. A, García-Medina, A. L, Galar-Martínez, M., Neri-Cruz, N., Islas-Flores, H., & Gómez-Oliván, L. M. (2013). The relationship of cytotoxic and genotoxic damage with blood aluminum levels and oxidative stress induced by this metal in common carp (*Cyprinus carpio*) erythrocytes. *Ecotoxicologia e Segurança Ambiental*, 96(1), 191-197. doi: 10.1016/j.ecoenv.2013.06.010
- Grisolia, C. K., Rivero, C. L. G., Starling, F. L. R. M., Silva, I. C. R., Barbosa, A. C., & Dorea, J. G. (2009). Profile of micronucleus frequencies and DNA damage in different species in different species of fish in a eutrophic tropical lake. *Genetics and Molecular Biology*, 32(1), 138-143. doi: 10.1590/S1415-47572009005000009
- Gensemer, R. W., & Playle, R. C. (1999). The bioavailability and toxicity of aluminium in aquatic environments. *Critical Reviews in Environmental Science and Technology*, 29(4), 315-450. doi: 10.1080/10643389991259245
- Guiloski, I. C., Ribas, J. L. C., Pereira, L. S., Perbiche, A. P. N., & Assis, H. C. S. (2015). Effects of trophic exposure to dexamethasone and diclofenac in freshwater fish. *Ecotoxicology and Environmental Safety*, 114(1), 204-211. doi: 10.1016/j.ecoenv.2014.11.020
- Helena, B. A., Vega, M., Barrado, E., Pardo, R., & Fernández, L. (1999). A case of hydrochemical characterization of alluvial aquifer influenced by human activities. *Water, Air and Soil Pollution*, 112(3), 365-387. doi: 10.1023/A:1005065422156
- Hemachandra, C. K., & Pathiratne, A. (2016). Combination of physico-chemical analysis, *Allium cepa* test system and *Oreochromis niloticus* erythrocyte based comet assay/nuclear abnormalities tests for cyto-genotoxicity assessments of treated effluents discharged from textile industries. *Ecotoxicology and environmental safety*, 131(1), 54-64. doi: 10.1016/j.ecoenv.2016.05.010
- Hoofman, R. N., & Raat, W. K. (1982). Induction of nuclear anomalies (micronuclei) in the peripheral blood erythrocytes of the eastern mudminnow *Umbra pygmaea* by ethyl methanesulphonate. *Mutation Research*, 104(1-3), 147-52. doi: 10.1016/0165-7992(82)90136-1
- Instituto Brasileiro de Geografia e Estatística [IBGE]. (2012). Indicadores de desenvolvimento sustentável: Brasil 2012. *Estudos & pesquisas. Informação geográfica*, Rio de Janeiro.
- Ide, A. H., Osawa, R. A., Marcante, L. O., Pereira, J. C., & Azevedo, J. C. R. (2017). Occurrence of pharmaceutical products, female sex hormones and caffeine in a subtropical region in Brazil. *CLEAN – Soil, Air, Water*, 45(9), 1-9. doi: 10.1002/clen.201700334
- Lieschke, G. J., & Currie, P. D. (2007). Animal models of human disease: zebrafish swim into view. *Nature Reviews Genetics*, 8(5), 353-367. doi: 10.1038/nrg2091
- Livingstone, D. R. (1998). The fate of organic xenobiotics in aquatic ecosystems: quantitative and qualitative differences in biotransformation by invertebrates and fish. *Comparative Biochemistry and Physiology*, 120(1), 43-49. doi: 10.1016/S1095-6433(98)10008-9
- Lobón-Cervía, J., Hess, L. L., Melack, J. M., & Araujo-Lima, C. A. R. M. (2015). The importance of forest cover for fish richness and abundance on the Amazon floodplain. *Hydrobiologia*, 750(1), 245-255. doi: 10.1007/s10750-014-2040-0
- Matozo, F., Turek, J. A., & Noleto, R. B. (2015). Avaliação dos efeitos genotóxicos do fungicida Ridomim em *Astyanax altiparanae* (Pisces, Characiformes). *Luminária*, 17(1), 121-131.
- Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'Hara, R. B., ... Wagner, H. (2016).

- vegan: community ecology package*. R package version, 2-0. Retrieved on Nov. 12, 2017 from <https://cran.r-project.org/web/packages/vegan/vegan.pdf>
- Pacheco, M., & Santos, M. A. (1998). Induction of liver EROD and erythrocytic nuclear abnormalities by cyclophosphamide and PAHs in *Anguilla anguilla* L. *Ecotoxicology and Environmental Safety*, 40(1), 71-76. doi: 10.1006/eesa.1998.1644
- Peterjohn, W. T., & Correll, D. L. (1984). Nutrient dynamics in an agricultural watershed: observations on the role of a riparian forest. *Ecology*, 65(5), 1466-1475. doi: 10.2307/1939127
- Ragugnetti, M., Adams, M. L., Guimarães, A. T. B., Sponchiado, G., Vasconcelos, E. C., & Oliveira, C. M. R. (2011). ibuprofen genotoxicity in aquatic environment: an experimental model using *Oreochromis niloticus*. *Water, Air, & Soil Pollution*, 218(1), 361-364. doi: 10.1007/s11270-010-0698-0
- Ribas, J. C., Zampronio, A. R., & Assis, H. C. S. (2015). Effects of trophic exposure to diclofenac and dexamethasone on hematological parameters and immune response in freshwater fish. *Environmental Toxicology and Chemistry*, 35(4), 975-982. doi: 10.1002/etc.3240
- Ribeiro, C. A. O., Katsumiti, A., França, P., Maschio, J., Zandoná, E., Cestari, M. M., ... Neto, F. F. (2013). Biomarkers responses in fish (*Atherinella brasiliensis*) of Paranaguá bay, southern Brazil, for assessment of pollutant effects. *Brazilian Journal of Oceanography*, 61(1), 1-11. doi: 10.1590/S1679-87592013000100001
- Seiler, R. L., Zugg, S. D., Thomas, J. M., & Howcroft, D. L. (1999). Caffeine and pharmaceuticals as indicators of waste water contamination in wells. *Ground Water*, 37(3) 405-410. doi: 10.1111/j.1745-6584.1999.tb01118.x
- Silva, M. R. L. R. (2014). Avaliação da toxicidade celular do herbicida glifosato em *Astyanax* spp. *Saúde e Meio Ambiente*, 3(2), 62-69. doi: 10.24302/sma.v3i2.629
- Silva, A. C., & Nepomuceno, J. C. (2010). Avaliação da frequência de micronúcleos em eritrócitos periféricos de mandi-amarelo (*Pimelodus maculatus*) do rio Paranaíba. *UNIPAM*, 1(7), 167-179.
- Sodré, F. F., Schnitzler, D. C., Scheffer, E. W. O., & Grassi, M. T. (2012). Evaluating copper behavior in urban surface waters under anthropic influence. A case study from the Iguaçu river, Brazil. *Aquatic Geochemistry*, 18(5), 389-405. doi: 10.1007/s10498-012-9162-7
- Tabacchi, E., Correll, D. L., Hauer, R., Pinay, G., Planty-Tabacchi, A. M., & Wismar, R. C. (1998). Development, maintenance and role of riparian vegetation in the river landscape. *Freshwater Biology*, 40(3), 497-516. doi: 10.1046/j.1365-2427.1998.00381.x
- Türkmen, M., Türkmen, A., Tepe, Y., Töre, Y., & Ates, A. (2008). Determination of metals in fish species from Aegean and Mediterranean seas. *Food Chemistry*, 113(1), 233-237. doi: 10.1016/j.foodchem.2008.06.071
- Zenkner, F. F., Soares, A. P. T., Prá, D., Köhler, A., & Rieger, A. (2011). Avaliação genotoxicológica em peixes nativos do Rio Pardinho, RS, Brasil. *Caderno de Pesquisa Série Biologia*, 23(1), 5-16. doi: 10.17058/cp.v23i1.4725

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